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Rising CO₂ from historical concentrations enhances the physiological
 performance of *Brassica napus* seedlings under optimal water supply but not
 under reduced water availability.

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11 Running title

12 Water stress at different [CO₂]

13 Keyword index

Canola, elevated CO₂, climate change, photosynthesis, stomatal conductance, shoot
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16 Abstract

The productivity of many important crops is significantly threatened by water shortage and the elevated atmospheric CO_2 can significantly interact with physiological processes and crop responses to drought. We examined the effects of three different CO_2 concentrations (historical ~300 ppm, ambient ~400 ppm and elevated ~700 ppm) on physiological traits of oilseed rape (*Brassica napus* L.) seedlings subjected to well-watered and reduced water availability. Our data show i)

that, as expected, increasing CO₂ level positively modulates leaf photosynthetic 23 traits, leaf water-use efficiency and growth under non-stressed conditions although a 24 pronounced acclimation of photosynthesis to elevated CO₂ occurred; ii) that the 25 predicted elevated CO₂ concentration does not reduce total evapotranspiration under 26 drought when compared to the present (400 ppm) and historical (300 ppm) 27 concentrations because of a larger leaf area that does not buffer transpiration; iii) that 28 accordingly the physiological traits analysed decreased similarly under stress for all 29 CO_2 concentrations. Our data support the hypothesis that increasing CO_2 30 31 concentrations may not significantly counteract the negative effect of increasing drought intensity on Brassica napus performance. 32

33 Introduction

The concentration of atmospheric CO₂ ([CO₂]) has been increasing steeply from 34 1958 (~316 ppm) to 2016 (>400 ppm) (NOAA Mauna Loa Observatory, Hawaii) and 35 36 the Intergovernmental Panel on Climate Change has predicted a [CO₂] in 2050 37 between ~550 and ~700 ppm. From an agricultural point of view, the elevated [CO₂] has been related to direct effects on crops' physiological processes (Leakey et al. 38 2012). Indeed, C₃ crops (Fitzgerald *et al.* 2016; O'Leary *et al.* 2015) and, to a lesser 39 extent C₄ crops (Ruiz-Vera *et al.* 2015), exhibited i) higher photosynthetic rates due 40 to an increased Rubisco carboxylation activity and a decreased oxygenation 41 reaction; ii) a lower stomatal conductance (g_s) following a depolarization of the 42 membrane potential of the guard cells accompanied by an increase of the outward 43 activity of K⁺ channels and a subsequent increase in guard cell Ca²⁺ concentration 44 (Leakey et al. 2012, Ainsworth & Rogers, 2007). These physiological changes have 45 been related to increasing biomass production and ameliorated responses of plants 46 47 to drought by an improvement of water-use efficiency at the canopy level (Leakey et

al. 2012). It is well-recognized that drought is the main factor affecting global crop 48 production and food security (Cattivelli et al. 2008) and climate change is predicted 49 to significantly increase the magnitude of water shortage and negatively affect the 50 yield of crops (Parmesan & Yohe, 2003). In this context, oilseed rape (OSR, 51 Brassica napus L.) is becoming one of the most popular oilseed crops in the world. 52 Drought periods over the most drought-sensitive phenological stages for OSR 53 (anthesis until mid-pod development) can lead up to 40% of seed yield losses (Faralli 54 et al. 2016). 55

56 It is therefore of pivotal importance to understand the physiological mechanisms that relate atmospheric CO₂ to drought and to determine whether the expected 57 detrimental effects on crops following the increases in extreme environmental 58 conditions could be at least mitigated by increases in CO₂ concentrations. A large 59 amount of work showed positive effects of elevated atmospheric CO₂ concentration 60 on crop's drought tolerance and hence yield reduction mitigation (Baker et al. 1997; 61 Wall 2001; Qaderi et al. 2006; O'Leary et al. 2015; Fitzgerald et al. 2016). However, 62 experiments on OSR have shown that elevated CO₂ concentrations may not mitigate 63 the detrimental effects of increasing temperatures on yield (Frenck et al. 2011). In 64 addition, Gray et al. (2016) have recently shown in multi-year FACE experiments that 65 the improvements in physiological performance and yield of soybean from elevated 66 67 [CO₂] can be counteracted by severe drought conditions. Therefore we conducted three experiments in the same controlled environment room at different CO2 68 concentrations (~300 ppm – historical 1950/1960 [CO₂], h[CO₂]; ~400 ppm – ambient 69 70 [CO₂], _a[CO₂]; ~700 ppm – elevated [CO₂], _e[CO₂]) and all three comparing the same combinations of watering regimes. The aims were to i) explore the physiological 71 72 effects of these [CO₂] on winter OSR seedlings growth and development: ii) to evaluate the interactions between the different [CO₂] and drought; iii) to characterize
and evaluate the relationships between physiological traits involved in the response
of OSR to different [CO₂] and drought.

76 Materials and methods

77 Plant materials and experimental layout

Winter oilseed rape (OSR, Brassica napus L.) seeds (cv. Excalibur, Dekalb, UK) 78 were sown on 8th January 2016 for Experiment I, 20th January 2016 for Experiment 79 II, and 9th February 2016 for Experiment III in individual pots (pot width 11 cm, pot 80 height 13 cm, total volume \sim 1500 cm³) containing the same amount (650 g + 25g of 81 pot weight) of a peat, sand, and pumice plant cultivation substrate (SoMi 513, 82 Dachstauden; Hawita, Vechta, Germany) characterized with a water retention curve 83 84 as described by Barboza-Barguero et al. (2015). Plants were germinated in a growth chamber at 20°C day / 15°C night, 12 / 12 photoperiod, 60 % RH and ambinet [CO₂]. 85 PAR was provided by neon lamps at ~200 μ mol m⁻² s⁻¹ at shoot apex measured with 86 a LI-190R PAR sensor (LI-COR, Lincoln, Nebraska USA). Plants were watered daily 87 with rain water to maintain the optimum soil moisture conditions for plant germination 88 and growth (around 90% of available water content, AWC, according to a water 89 retention curve, as described in Barboza-Barquero et al., 2015). Three seeds were 90 sown in each pot and for all the experiments two emerged seedlings were removed 91 after 4 days from emergence. At the 2nd leaf stage (GS 1.2 BBCH growth scale, 19th 92 January for Experiment I, 7th February for Experiment II and 25th February 2016 for 93 Experiment III) seedlings were placed in a controlled environment chamber (Jülich 94 95 Forschungszentrum, IBG-2 Plant Science, walk-in custom-made chamber) at 20°C day / 15°C night, 16/8 photoperiod, 60% RH and a VPD of 0.94 kPa. PAR was 96

97 provided with HPI-T-plus lamps (400W/645 E40, Philips, Netherlands) at an average 98 of ~500 μ mol m⁻² s⁻¹ at plant level with a minimum value of 460 μ mol m⁻² s⁻¹ and a 99 maximum of 540 μ mol m⁻² s⁻¹.

Each experiment consisted of five randomized blocks with two treatments (wellwatered - WW and droughted - WS) and six replicates in each block. Treatments were applied when plants reached the 4th leaf stage based on visual observations (GS 1.4, BBCH oilseed rape growth scale) (days after treatment 0, DAT 0 - 25th January for Experiment I, 12th February for Experiment II and 1st March 2016 for Experiment III).

According to the soil retention curve of the SoMi substrate, target weight at "pot capacity" (100% of available water content) was set to 1070 g of pot weight (including 25 g of pot weight and 20 g of plant fresh weight at the 4th leaf stage) and the permanent wilting point was set at 600 g. Thus the total AWC of the pots was accounted to be 425 ml. Hence, well-watered pots (WW, n=6) were weighed daily with a MC1 balance (Sartorius, Göttingen, Germany) and then rewatered to avoid any decrease in soil moisture according to the equation:

113

Eqn (1) Water to add = "pot capacity" weight – pot weight

Watering was withheld to water stressed pots (WS) over the whole experimental period (12 days). Pot saucers were used to avoid any water uptake. Total daily pot evapotranspiration (ET) was calculated as the water to add for WW and the daily reduction in pot weight for WS.

For each individual experiment, the chamber was set at a different $[CO_2]$: Experiment I 400 ppm, $_a[CO_2]$; Experiment II 700 ppm, $_e[CO_2]$; and Experiment III 300 ppm, $_h[CO_2]$. A respirator (Fig. S1) made up of a tube connected with a plastic mask on one side (respirator) was developed and used in the growth chamber to avoid

undesired [CO₂] fluctuations due to experimenter breathing during data collection and plant manipulation. The average [CO₂] for each experiment was 402.6 \pm 10.1 (SD) for Experiment I, 700.9 \pm 9.7 for Experiment II and 285.5 \pm 14.3 for Experiment III (Fig. S2)

126 Growth and relative water content analysis

On DAT 1, 4, 8 and 12, plants (n=6) from each treatment were harvested for all the 127 experiments. RWC was calculated according to Barr & Weatherley (1962) and was 128 determined as described by Faralli et al. (2015). Briefly, between 12:00 and 13:00, 129 one piece of leaf tissue of the fourth leaf (~2 cm²) was collected using a scalpel for 130 each plant (n=6). These samples were collected in a central region of the lamina 131 between the main vein and the leaf margin avoiding lateral veins. The fresh weight 132 was then recorded (Fw) and the samples were soaked in distilled water in Petri 133 dishes in the dark and at ~4°C during four hours. The turgid weight (T_w) was 134 recorded, the dried disks (oven-dried at 80°C for 24 hours) were weighed the day 135 after, and the dry weight was recorded (D_w). RWC (%) was then calculated 136 according to: 137

138 Eqn (2) RWC(%) =
$$\frac{F_W - D_W}{T_W - D_W} \times 100$$

After that, leaf number (LN) was counted and plants were excised at the base of the stem and leaf and stem fresh weight were recorded (Mettler Toledo XS 205 Dual Range, Columbus, USA). Total leaf area (LA) was recorded with a leaf area meter (LI-3100C area meter, Li-Cor, Lincoln, NE, USA). Samples were dried in an oven (Nabertherm P330, Nabertherm GmbH Lilienthal/Bremen, Germany) over 24 hours at 65°C and leaf, stem and total shoot dry weight were recorded. Specific leaf area was calculated as: SLA (g m⁻²) = leaf area / leaf DW.

Gas-exchange and chlorophyll fluorescence were simultaneously measured with a 147 Li-Cor 6400 (Li-Cor, Lincoln, NE, USA) with an integrated fluorescence leaf cuvette 148 (LI-6400-40; Li-Cor) on DAT 1, 4, 8 and 12 for all the experiments, between 09:00 149 and 12:00. To minimize potential leaf position and developmental age effects, all the 150 gas-exchange measurements were taken on the 4th fully expanded leaf of six 151 randomly selected plants for each treatment. In the Li-Cor cuvette, all the parameters 152 (leaf CO₂ assimilation at saturating light - A_{max} , stomatal conductance - g_s and the 153 ratio between sub-stomatal CO₂ and the ambient [CO₂] - C_i/C_a) were collected at 400 154 ppm CO₂ (Experiment I), 700 ppm CO₂ (Experiment II) and 300 ppm CO₂ 155 (Experiment III). Cuvette temperature was maintained at 25°C, photosynthetically 156 active photon flux density (PPFD) was 1200 µmol m⁻² s⁻¹ (saturating PPFD for OSR 157 seedlings, previously evaluated by light curves, Fig. S3), with a 10:90 blue:red light 158 and a flow rate of 300 µmol s⁻¹. Intrinsic water-use efficiency was calculated as 159 A_{max}/q_s . The ratio (A_{max}/q_s) is considered to be more reliable than the water-use 160 efficiency calculated from CO₂ assimilation to transpiration rate, since it is not 161 affected by the VPD inside the leaf chamber (Webster et al. 2016). The data were 162 163 collected after the leaf achieved a steady-state. The data were collected after the leaf achieved a steady-state, and minimum fluorescence (F_s) and maximum 164 fluorescence (F_m) were recorded by ensuring a light-saturating pulse of 8000 µmol 165 m⁻² s⁻¹. The actual photochemical efficiency of the photosystem II ($\Delta F/F_m$) was 166 calculated according to Baker (2008) as follow: 167

168 Eqn (3) $\Delta F/Fm' = \frac{Fm'-Fs}{Fm'}$

and the electron transport rate as:

Eqn (4)
$$J = \Delta F / Fm' \times PPFD \times \alpha \times \beta$$

171 Statistical analysis

Available water content and daily evapotranspiration data from the three experiments 172 are daily means (n=6) ± standard error (SE). Carbon dioxide treatments are means 173 $(n=1440) \pm \text{standard deviation (SD)}$. With the exception of $[CO_2]$, the parameters 174 inside the growth chamber (i.e. RH, temperature, light and VPD) were very similar or 175 identical during the three experiments. Also, since the growth conditions of the three 176 experiments were very similar and no morphological differences were recorded 177 between the three sets of plant material, a hypothetical time-specific effect affecting 178 the CO₂ treatments was not considered a possibility. Therefore all the physiological 179 traits were analysed with a two-way ANOVA (CO₂ x watering regimes, Figure S4) in 180 GenStat 17th Edition (VSN International, UK). However, since the three experiments 181 were formally independent, the homogeneity of variance for each two-way ANOVA 182 was checked by visually examining the distribution histogram of the residuals and 183 the fitted vs. residuals plot. While the distribution was acceptable for all the traits and 184 185 DAT, in five cases (RWC DAT 12, Leaf Area DAT 8, Amax DAT 12, A_{max}/g_s DAT 1) the fitted vs. residuals plot showed heteroscedasticity. These data were transformed 186 to a natural logarithmic scale and in all cases the heteroscedasticity was reduced 187 and the distribution was improved. Subsequently the log-transformed data were 188 subjected to two-way ANOVA and the P-values for all the interactions were 189 recorded. In all the cases logarithmic transformation had a negligible effect on P-190 values suggesting that the data heteroscedasticity was not significantly affecting the 191 data. Therefore, due to the above evidence, the data are statistically analysed and 192 193 presented in their original form without logarithmic transformation and thus including

[CO₂] factor in the ANOVA model. The two-way ANOVA outputs are presented inSupplementary Fig. 4.

196 **Results**

197 Total plant evapotranspiration and pot available water content

Total plant ET of WW was similar at each [CO₂], increasing from ~50 mL at DAT 1 to 198 ~130 mL at DAT 7 (Fig. 1A). From DAT 8, while h[CO₂] maintained total ET at ~ 180 199 mL, the plants grown under a[CO₂], slightly decreased the ET from ~160 mL to ~120 200 mL. On the contrary, _e[CO₂], steadily maintained a significant lower ET value from 201 ~100 ml at DAT 8 to ~60 mL at DAT 12. Therefore, pot AWC of WW plants from DAT 202 8 to DAT 12 was higher in _e[CO₂] conditions than under _a[CO₂] and, to a greater 203 extent, h[CO₂] (Fig. 1B). WS treatment decreased both ET and AWC. Under WS 204 205 conditions, e[CO₂] plants showed a significantly higher ET when compared to a[CO₂] and h[CO₂] but only until DAT4 (Fig. 1C). From DAT 5, similar decreases in ET were 206 recorded between [CO₂]. Therefore, pot AWC of WS plants was lower under e[CO₂] 207 than under a[CO2] and h[CO2], although PWP was reached similarly for all the [CO2] 208 between DAT 7 and DAT 8 (Fig. 1D). 209

210 Growth analysis

Leaf number (LN) significantly increased over time in WW plants grown at all the [CO₂] (from 4 leaves at DAT 1 to 7.5-8 leaves at DAT 12) and no significant differences were found between the different [CO₂] treatments (Fig. 2A). WS significantly decreased LN compared to the WW plants when grown at h[CO₂] and a[CO₂] since DAT 4 (Fig. 2B). However e[CO₂] showed a reduction in LN under WS only from DAT 8.

Over the twelve days of the experiments, WW plants grown in a[CO₂] had a shoot 217 DW (Fig. 2C) that was significantly higher than that of the plants grown in $h[CO_2]$ by 218 34% on average (Fig. 2C) (P<0.001 for DAT 4, 8 and 12). Increases in shoot DW of 219 20% on average were observed in plants grown at e[CO₂] (Fig. 2C) with respect to 220 the plants grown under a[CO₂]. Significant reductions in shoot DW were observed 221 under WS on DAT 8 and DAT 12 at all the [CO₂] when compared to the WW plants 222 (Fig. 2D). However, WS plants grown under _e[CO₂] exhibited a higher DW compared 223 to h[CO₂] and a[CO₂] by 42% and 33% on DAT 4, 40% and 26% on DAT 8 and 37% 224 225 and 27% on DAT 12, respectively .

Leaf area (LA) of WW plants grown under a[CO₂] significantly increased with time 226 from 190 cm² on DAT 1 to 800 cm² on DAT 12 (Fig. 2E). Under h[CO₂], WW plants 227 exhibited a similar significant increase in LA that was however lower than that of the 228 plants grown at _a[CO₂] (Fig. 2E) (from 190 cm² on DAT 1 to 590 cm² on DAT 12). 229 _e[CO₂] stimulated LA of WW plants leading to an average LA of 1190 cm² on DAT 12 230 (Fig. 2E). For DAT 4 and DAT 8, however, no significant differences in LA were 231 found between plants grown at e[CO₂] and a[CO₂]. WS had a significant effect on LA 232 at all the [CO₂] and the decrease was significant since DAT 4 in _e[CO₂], _a[CO₂] and 233 h[CO₂]-grown plants (Fig. 2F). 234

Specific leaf area (SLA) decreased over time on WW plants grown at all the $[CO_2]$ and WS significantly decreased SLA values at all the $[CO_2]$ (P<0.001 since DAT 4) (Fig. 2G and H).

238 Plant water status

The leaf RWC of WW plants fluctuated from 90% to 91% on average and no significant differences were observed between the three $[CO_2]$ (Table 1). Under WS conditions, leaf RWC significantly decreased from DAT 4 at all the $[CO_2]$ (*P*<0.001 at

all the DAT). RWC of $_{e}[CO_{2}]$ plants under WS conditions was significantly higher (RWC 70%, *P*=0.016) than $_{a}[CO_{2}]$ (RWC 62%) and $_{h}[CO_{2}]$ (RWC 67%) at DAT 8. However, since the drop in RWC was similar for all the other DAT, RWC values were not overall significantly different under WS between [CO₂] leading to very similar values at DAT 12.

247 Gas-exchange

WW plants grown under $_{a}[CO_{2}]$ showed an average A_{max} of 26 µmol m⁻² s⁻¹ that was 229 22% lower than that of the plants grown in $_{e}[CO_{2}]$ (Fig. 3A). On the contrary, $_{a}[CO_{2}]$ 250 exhibited an increase in A_{max} by 12% on average with respect to the $_{h}[CO_{2}]$ (Fig. 3A). 251 Plants under WS conditions exhibited a reduction in A_{max} compared to the WW 252 plants that was significant from DAT 4 for all the [CO₂] (*P*<0.001) (Fig. 3B).

WW plants grown under $_{a}[CO_{2}]$ exhibited a g_{s} of 0.67 mol mm⁻² s⁻¹ on average, showing a decreasing slope over the last days of the experiment (Fig. 3C). With respect to the $_{a}[CO_{2}]$, a 7% increase on average in g_{s} was observed in $_{h}[CO_{2}]$ -grown plants (Fig. 3C), without showing any g_{s} reduction over time. On the contrary, g_{s} was significantly reduced over time in $_{e}[CO_{2}]$ plants and by 40% on average with respect to the $_{a}[CO_{2}]$ (Fig. 3C). WS conditions reduced g_{s} by 64%, 55% and 40% under $_{h}[CO_{2}]$, $_{a}[CO_{2}]$ and $_{e}[CO_{2}]$ respectively (Fig. 3D).

WW plants grown under $_{h}[CO_{2}]$ and $_{a}[CO_{2}]$ showed a relatively stable A_{max}/g_{s} over time with an average value of 31 µmol mol⁻¹ and 40 µmol mol⁻¹ respectively (Fig. 3E). On the contrary, $_{e}[CO_{2}]$ -grown plants showed a significant increase in time from DAT 1 to DAT 12 leading to an average value of 90 µmol mol⁻¹ (Fig. 3E). WS plants had higher A_{max}/g_{s} values with respect to the WW ones, with an average of 52, 76 and 95 µmol mol⁻¹ for $_{h}[CO_{2}]$, $_{a}[CO_{2}]$ and $_{e}[CO_{2}]$ -grown plants respectively (Fig. 3F). Under WW conditions, ETR values of $_{e}[CO_{2}]$ -grown plants was on average higher 267 (190 μ mol e⁻¹ m⁻² s⁻¹) than $_{h}[CO_{2}]$ and $_{a}[CO_{2}]$ (177 and 182 190 μ mol e⁻¹ m⁻² s⁻¹ 268 respectively) until DAT 8. Conversely, on DAT 12, similar ETR values were recorded 269 between $_{h}[CO_{2}]$ and $_{a}[CO_{2}]$ (171 and 164 μ mol e⁻¹ m⁻² s⁻¹ respectively) that were 270 lower compared to the $_{e}[CO_{2}]$ (154 μ mol e⁻¹ m⁻² s⁻¹). Under WS, declines in ETR 271 values were evident for all the $[CO_{2}]$ and although similar values were recorded at 272 DAT 12, the reduction was less pronounced under $_{e}[CO_{2}]$ at DAT 8.

273 **Discussion**

[CO₂] influenced biomass accumulation and leaf gas-exchange under WW

275 conditions

In our experiments, e[CO₂] at 700 ppm stimulated Amax by ~22% (in line with the 276 findings of Qaderi et al. (2006)) and h[CO2] reduced Amax by ~12% when both 277 compared to a[CO₂] under WW conditions. The increase under e[CO₂] found in OSR 278 seedlings was however lower than in other C_3 crops such as durum wheat (~37%; 279 Aranjuelo et al. 2015) and rice (~40%, Vu et al. 1997) and in our experiments at DAT 280 12 the A_{max} of e[CO₂] plants was similar to that of a[CO₂]. This suggests strong 281 photosynthetic acclimation potentially due to the increased LA of plants at e[CO2] that 282 283 lead to a reduction in Rubisco activity due to a depression in area-based leaf N content. In addition, the concomitant decrease in ETR and Amax that occurred under 284 $_{e}[CO_{2}]$ but not under $_{h}[CO_{2}]$ and $_{a}[CO_{2}]$ at DAT 12 can be also an indication of 285 signals derived from increases in sucrose transport to guard cells (Outlaw, 2003), 286 thylakoid membrane damage (Haworth et al. 2016) and/or increase in soluble sugar 287 in the apoplast that reduced the mesophyll signal (Haworth et al. 2016). 288

The initial enhanced A_{max} following the increasing [CO₂] was accompanied by larger LA and shoot DW, consistent with previous work in OSR (Qaderi *et al.* 2006). This

report however is the first showing differences in A_{max}, LA and shoot DW between 291 _a[CO₂] and _h[CO₂]-grown OSR. It is not clear whether increased A_{max} following high 292 [CO₂] can enhance sink activity but carbohydrate translocation can be reduced when 293 sink size is limited (Arp, 1991). A FACE experiment with spring canola grown at 500 294 ppm [CO₂] showed no significant increases in yield compared to the ambient 295 concentration-grown plots (Franzaring et al. 2008) probably due to limited carbon 296 storage structures, therefore a potential small sink size (Reekie et al. 1998). Our 297 data however indicate that OSR was potentially source-limited in the first years of its 298 299 agricultural production (~1960) and that rising [CO₂] may have played a role at boosting the steady increase in OSR seed yield seen in recent years (Berry and 300 Spink, 2006). 301

It has been proposed that the larger plant size that can be achieved at elevated CO₂ 302 303 is related to higher water use (hence transpiration) (Xu et al. 2013; Polley et al. 2008; Nowak et al. 2004). Our results show that in OSR seedlings grown under WW 304 conditions the standardized transpiration over time for LA is increased under a[CO₂] 305 $(1106 \pm 105 \text{ mmol } H_2O)$ when compared to the $h[CO_2]$ (713 ± 105 mmol $H_2O)$, but 306 not under $_{e}[CO_{2}]$ (852 ± 105 mmol H₂O) suggesting that the short-term acclimation to 307 308 _e[CO₂] is highly beneficial in the WW plants even if the LA is dramatically increased. In addition, ET data show that from DAT 8, e[CO₂] plants are subjected to a 309 significant reduction in water loss, potentially due to the larger leaf area that 310 attenuates soil evaporation. This confirms that, although plant water loss and LA are 311 not linearly correlated, larger LA can buffer the greater areas devoted to transpiration 312 when self-shading and/or elevated leaf-area index threshold are reached under non-313 stressed conditions. 314

[CO₂] did not affect the whole-plant response under WS conditions.

316 In our experiments, RWC of WS plants was on average 89, 80, 66 and 54%, that reflects a good range of dehydration. When RWC was decreased to 66% (DAT 8), gs 317 was completely inhibited at all the [CO₂], in line with previous findings (Albert et al. 318 2012). These data suggest application of mild stress at DAT 4, severe stress at DAT 319 8 and terminal stress at DAT 12. We hypothesized that by maintaining low g_s , 320 elevated atmospheric [CO₂] may ameliorate plant water status under stress by 321 preserving soil water content (Leakey et al. 2012). Polley et al. (1999) suggested that 322 the effects of elevated [CO₂] on osmotic adjustment are minimal, indicating that the 323 324 maintenance of high plant water status in plants grown at elevated [CO₂] under stress is a result of stomatal control. However, and contrary to expectation, in our 325 experiments no differences were found in RWC between [CO₂] treatments. In our 326 327 experiments the larger leaf area produced under _e[CO₂] led to a similar transpiration rate standardized for the LA (~200 mmol m⁻²s⁻¹ for all the [CO₂] under stress) thus 328 offsetting the water-loss reduction following the e[CO₂]-induced stomatal closure. 329 Recently Grav et al. (2016) showed that, at field level, elevated atmospheric CO₂ 330 concentration (from 550 to 585 ppm) does not always conserve water in soil 331 following reduction in transpiration rate. This leads to non-significant benefits in 332 terms of soybean yield when drought progressively increases. Since the relation 333 between the increase in LA and transpiration in the field is not linear due to the self-334 335 shading effect that occur when large LA are reached, Gray et al. (2016) hypothesised that in wet years (large baseline LA), higher atmospheric [CO₂] may 336 significantly reduce crop WU, consistently with our WW data on OSR (as explained 337 above). However this may not occur in dry years, where increases in LA may only 338 lead to higher plant water loss, similarly to our findings on WS. To confirm that, in our 339 experiments A_{max} -to- g_s curves show that in the first part of soil drying, g_s is reduced 340

more than A_{max} leading to higher A_{max}/g_s values for all the [CO₂] treatments (Fig. 4A). 341 However, despite the curve plateau being higher at e[CO₂], the shape of the A_{max}-to-342 g_s curves were similar for all the [CO₂], possibly because the increased LA resulted 343 in similar detrimental effects on A_{max} and RWC (Fig. 4B), thus similarly reducing g_s 344 (Fig. 4C), between the [CO₂] under stress. This hypothesis is confirmed by ET data 345 where e[CO₂] plants exhibited a very high ET from DAT 1 to DAT 4 due to higher LA 346 and LN that lead to fast use of the water resources available. Therefore, although 347 leaf traits collected with gas-exchange system (e.g A_{max} and A_{max}/g_s) were higher on 348 349 DAT 4 (therefore under a moderate stress) under e[CO₂], increasing drought intensity leads to similar detrimental effects on OSR physiology regardless of the [CO₂] at 350 which they are grown. 351

352 Conclusions

The work presented here shows improvements in the physiological performance of 353 winter OSR seedlings in elevated [CO₂], mainly by stimulating photosynthesis and 354 355 thus increasing leaf area in line with work focusing in other major food crops. Strong differences were found between plants grown at 300 and 400 ppm suggesting that 356 OSR may have been significantly source-limited in the first years of its agricultural 357 production (~1960). Increasing the [CO₂] decreased leaf stomatal conductance and 358 improve leaf water-use efficiency but no significant beneficial effects were found 359 under drought due to the larger leaf area gained over the first few days of stress 360 (when water was available) that offsets the elevated [CO₂]-induced stomatal closure 361 leading to similar total ET between the [CO₂] treatments. Our data are in line with 362 recent findings in field-grown soybean, and shows this behaviour on another crop 363 species and over a much broader range of [CO₂]. 364

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Figures and tables



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Figure 1. Development of daily total plant evapotranspiration (ET, mL) under wellwatered (WW, A) and droughted conditions (WS, C) and development of pot available water content (AWC, mL) under well-watered (WW, B) and droughted conditions (WS, D) of winter oilseed rape plants grown at historical (300 ppm), ambient (400 ppm) and elevated (700 ppm) atmospheric CO₂ concentration. DAT= days after treatment application. Data were collected daily with a balance. Data are means (n=6) ± SE.



Figure 2. Development of leaf number (A, B), shoot dry weight (C, D; DW, g), total
leaf area (E, F; LA, cm² plant⁻¹) and specific leaf area (G, H; SLA cm² g⁻¹) of winter
oilseed rape plants grown at historical (300 ppm), ambient (400 ppm) and elevated
(700 ppm) atmospheric CO₂ concentration under well-watered (WW; A, C, E, G) and
water-stress (WS; B, D, F, H) conditions. Data were collected at days after treatment
application (DAT) 1, 4, 8 and 12. Data are means



Figure 3. Development of carbon assimilation rate at saturating light (A,B; A_{max}), stomatal conductance (C, D; g_s), intrinsic water-use efficiency (E, F; A_{max}/ g_s) and electron transport rate (G, H; ETR) of winter oilseed rape plants grown at historical (300 ppm), ambient (400 ppm) and elevated (700 ppm) atmospheric CO₂ concentration under well-watered (WW; A, C, E) and water-stress (WS; B, D, F) conditions. Data were collected at days after treatment application (DAT) 1, 4, 8 and 12. Data are means (n=6) ± SE.





Figure 4. Correlations between A) CO₂ assimilation rate at saturating light (A_{max}) and stomatal conductance (g_s) ,B) A_{max} and leaf relative water content (RWC) and C) g_s and RWC, of winter oilseed rape plants grown at 300 ppm (black symbols), 400 ppm (grey symbols) and 700 ppm white symbols) atmospheric CO₂ concentration under well-watered (WW) and water-stress (WS) conditions Data points are means (n=6). Lines were fitted using exponential curve for A, linear regression for B and polynomial regression for C.

	WW	WS					
[CO ₂]	RWC (%	6) DAT 1	ANOVA Factor	P-values			
300 ppm	89.9	88.9	CO ₂	0.064			
400 ppm	89.0	89.0	Watering regimes	0.802			
700 ppm	90.1	90.0	CO2 x Watering regimes	0.869			
RWC (%) DAT 4							
300 ppm	89.5	82.7	CO ₂	0.182			
400 ppm	90.2	76.6	Watering regimes	<.001*			
700 ppm	90.8	81.6	CO2 x Watering regimes	0.120			
RWC (%) DAT 8							
300 ppm	88.7	67.7	CO ₂	0.016*			
400 ppm	90.9	61.9	Watering regimes	<.001*			
700 ppm	91.8	69.2	CO2 x Watering regimes	0.018*			
RWC (%) DAT 12							
300 ppm	89.9	56.2	CO ₂	0.501			
400 ppm	89.6	55.6	Watering regimes	<.001*			
700 ppm	91.8	52.3	CO2 x Watering regimes	0.065			

531 532 533 534 535	Table 1. Relative water content data (RWC,%) of winter oilseed rape plants grown at historical (300 ppm), ambient (400 ppm) and elevated (700 ppm) atmospheric CO_2 concentration under well-watered (WW) and water-stress (WS) conditions. Data were collected at days after treatment application (DAT) 1, 4, 8 and 12. Data are means (n=6) ± SE.
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Supplementary Material



Supplementary Figure 1. Example of experimental room with the bespoke respirator and the cardboard box used to cover plants before the light curve protocol.



560 Supplementary Figure 2. Development of atmospheric $[CO_2]$ over Experiment I 561 $(a[CO_2])$, Experiment II $(e[CO_2])$ and Experiment II $(h[CO_2])$. Data points are means 562 $(n=1440) \pm \text{standard deviation (SD)}$.



Supplementary Figure 3. Light-response curves for oilseed rape seedlings. The curves were plotted on the third fully expanded leaf before Experiment I. A Li-Cor 6400 was used supplemented with a LI-6400-40 cuvette. CO₂ assimilation rates were collected at 100, 200, 300, 500, 600, 800, 1000, 1200, 1500 and 2000 PPFD. Cuvette temperature was maintained at 25°C, the flow rate was 300 µmol s⁻¹, CO₂ (Ca) was 400 ppm and the light source was maintained at 10:90 blue-red light. Data points are means \pm SEM (*n*=6) with the exception of 200, 500 and 800 PPFD where n=3.

Supplementary Figure 4. Analysis of variance (*P*-values) for $[CO_2]$, watering regimes and their interactive effects on plant dry-weight (shoot DW), Leaf area, Leaf number, Specific Leaf Area, CO₂ assimilation rate at saturating light (A_{max}), stomatal conductance (*g*_s) and water use efficiency (A_{max}/*g*_s) of winter oilseed rape plants grown at 300ppm, 400ppm and 700ppm atmospheric CO₂ concentration under well-watered (WW) and water-stress (WS) conditions. Values significance is highlighted with asterisks (*P*<0.05)

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	Shoot DW					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	P-values					
CO ₂	0.219	<.001*	<.001*	<.001*		
Watering regimes	0.662	0.053	<.001*	<.001*		
CO ₂ x Watering regimes	0.829	0.358	<.001*	<.001*		
	Leaf Area					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	P-values					
CO ₂	0.333	<.001*	<.001*	<.001*		
Watering regimes	0.367	<.001*	<.001*	<.001*		
CO ₂ x Watering regimes	0.953	0.052	0.001*	<.001*		
	Leaf number					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	<i>P</i> -values					
CO ₂	0.232	<.001	<.001*	<.001*		
Watering regimes	1	0.016*	<.001*	<.001*		
CO2 x Watering regimes	0.929	0.097	0.005*	0.066		
	Specific Leaf Area					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	<i>P</i> -values					
CO ₂	0.15	<.001*	<.001*	<.001*		
Watering regimes	0.302	0.003*	<.001*	<.001*		
CO ₂ x Watering regimes	0.682	0.277	<.001*	0.45		
	A _{max}					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	P-values					
CO ₂	<.001*	<.001*	<.001*	0.008*		
Watering regimes	0.445	<.001*	<.001*	<.001*		
CO ₂ x Watering regimes	0.043*	0.063	<.001*	0.071		
	g₅					
	DAT 1	DAT 4	DAT 8	DAT 12		
ANOVA Factor	<i>P</i> -values					
CO ₂	<.001*	<.001*	<.001*	<.001*		
Watering regimes	0.254	<.001*	<.001*	<.001*		
CO2 x Watering regimes	0.71	<.001*	<.001*	<.001*		
		Am	_{ax} /g _s			
	DAT 1 DAT 4 DAT 8 DAT 12					
ANOVA Factor	<i>P</i> -values					
CO ₂	<.001*	<.001*	<.001*	<.001*		
Watering regimes	0.505	<.001*	<.001*	0.956		
CO ₂ x Watering regimes	0.536	0.062	0.003*	<.001*		