

Effect of rheological and structural properties of bacterial cellulose fibrils and whey protein biocomposites on electrosprayed food-grade particles

by Paximada, P., Kanavou, E. and Mandala, I.G.

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Corresponding Author: Dr. Ioanna Mandala, PhD

Corresponding Author's Institution: Agricultural University of Athens

First Author: Parskevi Paximada

Order of Authors: Parskevi Paximada; Eugenia Kanavou; Ioanna Mandala, PhD

Abstract: In this work, we investigated the role of bacterial cellulose nano-fibrils (BCNFs) as an alternative polymer to obtain food-grade particles with the electrospraying technique. Suspensions were prepared using BCNFs (1-16% wt) and whey protein isolate (WPI) in various concentrations (10-30% wt). Surface tension and electrical conductivity depended on the BC concentration and further increased by its increasing amount. A great increase in interfacial viscosity was also noticed according to the BCNFs concentration. A strong impact of BCNFs at the interface, influencing charge density and interactions of the two polymers was suggested. Different groups of the suspensions can be found that resulted in spherical nano- or submicron- particles by electrospraying. Uniform, nano-particles can be successfully produced taking into account the interfacial viscosity of the initial suspensions. Interfacial, compared to bulk viscosity, is a valuable tool to find out the appropriate suspension rheological properties in order to produce fine particles.

Research Data Related to this Submission

There are no linked research data sets for this submission. The following reason is given:

Data will be made available on request

Highlights

- Spherical nano- or submicron- particles obtained through electrospraying
- Surface tension and viscosity capable to predict the structure of BC-WPI particles
- Interfacial viscosity of the suspensions related to the particle size and uniformity

**1 Effect of rheological and structural properties of bacterial cellulose fibrils and whey
2 protein biocomposites on electrosprayed food-grade particles**

3 Paraskevi Paximada^a, Eugenia Kanavou^a, Ioanna. G. Mandala^{a1}

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5 ^a Food Science & Nutrition, Agricultural University of Athens, Iera Odos 75, 11855, Athens,
6 Greece, imandala@aua.gr

7

¹Corresponding author. Tel.: +30 2105294692
E-mail address: imandala@aua.gr (Ioanna Mandala)

1 **Abstract**

2 In this work, we investigated the role of bacterial cellulose nano-fibrils (BCNFs) as an
3 alternative polymer to obtain food-grade particles with the electrospraying technique.
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5 various concentrations (10-30% wt). Surface tension and electrical conductivity depended on
6 the BC concentration and further increased by its increasing amount. A great increase in
7 interfacial viscosity was also noticed according to the BCNFs concentration. A strong impact
8 of BCNFs at the interface, influencing charge density and interactions of the two polymers
9 was suggested. Different groups of the suspensions can be found that resulted in spherical
10 nano- or submicron- particles by electrospraying. Uniform, nano-particles can be successfully
11 produced taking into account the interfacial viscosity of the initial suspensions. Interfacial,
12 compared to bulk viscosity, is a valuable tool to find out the appropriate suspension
13 rheological properties in order to produce fine particles.

1 **1. Introduction**

2 Cellulose is a natural polysaccharide as it is the primary component of plant cell walls.
3 Because of its fibrous structure and physical strength, it has high applicability in material
4 industry e.g. paper and textile applications. In addition, bacterial cellulose (BC) is a natural
5 polysaccharide, produced by bacteria. BC has remarkable properties compared to plant-
6 based cellulose. The fibres of BC have higher aspect ratio, they are thinner, occupy larger
7 area than plant-based cellulose fibres, and can absorb a great water amount creating a stiff
8 3D network. (Nakagaito, Iwamoto & Yano, 2005; Nishi et al., 1990).

9 Commonly used hydrocolloids as thickeners are xanthan gum, locust bean gum, and
10 hydroxymethylcellulose. Our previous studies concluded that BC shows a greater shear
11 thinning behaviour than commercial thickeners (Paximada, Koutinas, Scholten & Mandala,
12 2016). BC is more efficient to increase the zero-shear viscosity and the rheological profiles of
13 BC-stabilized emulsions, meaning that less amount of BC is required to obtain the same
14 thickening-effect in a solution. Moreover, BC presents an exceptional bioaffinity resulting in
15 several applications though it is still produced in lab scale (Choi & Sin, 2020). Taking all the
16 above into account, we can summarize that BC is a cheaper alternative thickener.

17 Lately, protein and BCNF interactions have gained more attention in biocomposite production.
18 For example, BCNFs and hydrolysed soy protein result in improved rheological properties,
19 and cellulose fibrils enhance the solid-like behaviour of the dispersions (Gouzy, Tsekou,
20 Remilin & Velikov, 2019). Moreover, whey or soy protein isolates and BCNFs interact at the
21 interface of an emulsion, stabilising the droplets. The investigated interfacial and bulk
22 rheological properties give greater stability to the emulsion (Li et al., 2019; Paximada, Dubey
23 & Howarth, 2018). The complexes at the oil-water interface are formed due to the change of
24 cellulose wettability. This wettability is caused by change of the adsorption capacity of the
25 protein and polysaccharides.

26 Biopolymer particles may also be used to protect and deliver bioactive compounds in food
27 systems. Therefore, there has been considerable interest in the formation of biopolymer
28 nano-particles from proteins and/or polysaccharides. The most common techniques are
29 spray-drying (Zhang et al., 2019), emulsification or coacervation (Koupantsis et., 2016).
30 However, these techniques have significant drawbacks, such as the heating required for the

1 process and the residual traces of solvents. To mitigate these drawbacks,
2 electrohydrodynamic (EHD) processing and specifically electrospraying constitutes an
3 innovative technology that could replace the commonly used techniques. In the
4 electrospraying process, a high voltage is applied on a polymer solution drying it to particles
5 that are then used in many applications (Bhardwaj & Kundu, 2010). What is more, during
6 electrospraying the particles are charged, hence they repulse each other leading to the
7 elimination of particle agglomeration.

8 Electrospraying has distinctive advantages over conventional mechanical spraying systems
9 such as the smaller droplet size with narrower distribution, higher stability of the bioactives.
10 Parameters that influence the process are: polymer concentration, molecular weight and its
11 distribution, solvent quality and volatility, physical properties (surface tension, conductivity,
12 viscosity, viscoelasticity), and the equipment processing conditions (flow rate, distance
13 between the electrodes and applied potential difference). These parameters have been
14 analysed in many studies (Casper, Stephens, Tassi, Chase & Rabolt, 2004, Regev,
15 Vandebril, Zussman & Clasen, 2010, Theron, Zussman & Yarin, 2004).

16 Recently, proteins and polysaccharides have attracted significant attention in the search of
17 suitable encapsulants to be electrosprayed. However, pure proteins are difficult or even
18 impossible to be electrosprayed due to their three-dimensional network (López-Rubio and
19 Lagaron, 2012). These limitations can be overcome by blending them with polysaccharides,
20 such as starch (López-Rubio et al., 2012) or pullulan (Aceituno-Medina et al., 2013), which
21 significantly improve the natural polymer's sprayability. Additionally, new polymers are
22 particularly interesting molecules for electrospraying.

23 Hence, this study focuses on the properties of bacterial cellulose and whey protein isolate
24 mixtures in terms of their ability to be electrosprayed and to make particles for food
25 applications. The properties of the produced particles are also within the scope of this study.

26

27 2. Materials and methods

28 2.1 Materials

1 Whey protein isolate (WPI), Lactodan DI-9224, was kindly provided by Arla (Arla Foods
2 Ingredients, Amba-Denmark). Tween 20 was purchased from Fisher Scientific (St. Louis,
3 USA). All the other chemicals used were of analytical grade.

4 Bacterial cellulose (BC) was produced as described in our previous work (Paximada,
5 Koutinas, Scholten & Mandala, 2016b). Briefly, bacterial cultivations (*Komagataeibacter*
6 *sucrofermentans* DSM 15973) were carried out using a synthetic medium. Acid hydrolysis in a
7 cellulose/acid ratio of approximately 20g/L took place in BC by adding sulfuric acid (60% w/w,
8 4 days, 40°C) until homogenous suspension was obtained. BCNF were produced as a white
9 precipitate after several centrifugation (12.500 rpm) and washing cycles. The mean length of
10 untreated cellulose fibrils was up to 37.5±6.8 µm and their width up to 120±8 nm. After acid
11 treatment their length was 900±100 nm and their width 33±15 nm. It should be mentioned that
12 the differences between nanocrystals (BCNCs) and BCNFs, according to their length, can be
13 slight. Lengths of 100–1500 nm are referred to crystals (Choi & Sin, 2020), but we assume
14 that the length of about 1 µm, is long enough to consider that nanofibrils are produced.
15 Preliminary experiments showed that the type of acid, HCl or H₂SO₄, strongly affected the
16 morphology of the produced fibrils. By using H₂SO₄ the smallest fibril size was achieved.

17 *2.2 Preparation of the suspensions*

18 BC-WPI suspensions were prepared by dissolving a precalculated amount of WPI powder
19 and Tween 20 in the respective amount of BC suspension through gentle stirring for 2 hours
20 at room temperature. The pH of the suspensions was adjusted at 3.6. The resulting
21 suspensions reached a final concentration of 0- 16% wt BC, 10- 30% wt WPI, and 2% wt
22 Tween 20.

23 *2.3. Electro spraying process*

24 For this process, an electro spraying set-up (Fluidnatek LE-10, BioInicia S.L., Valencia, Spain)
25 was used. This set up included a high voltage power supply. The suspensions were
26 introduced in 5 mL syringes which were driven by a pump towards a stainless-steel needle
27 (inner diameter 0.7 mm). The needle was horizontal with respect to a grounded collector. The
28 experiment was carried out at ambient conditions. The electro spraying conditions were fixed
29 at 20-80 µL/h flow rate, 10-20 kV voltage and a tip-to-collector distance of 10 cm.

1 *2.4 Characterization of the electro sprayed suspensions*

2 The surface tension and conductivity of the WPI-BCNFS suspensions were measured using a
3 tensiometer (KSV Sigma 701, KSV Instruments Ltd., Finland) and a conductivity meter
4 (SensoDirect 110, Lovibond, Dortmund, Germany), respectively. ζ -potential measurements
5 were carried with Dynamic Laser Light Scattering (Zetasizer Nano ZS, Malvern Instruments,
6 Worcestershire, UK) at 25°C using the method described previously (Paximada, Koutinas,
7 Scholten & Mandala, 2016).

8 *2.5 Rheological properties of the electro sprayed suspensions*

9 Rheological measurements of the suspensions were performed on a stress-controlled
10 rheometer (Discovery HR-3, TA Instruments, New Castle, DE, USA) equipped with a
11 concentric cylinders geometry (30mm cup diameter, 28mm bob diameter) as described
12 previously (Paximada, et al., 2016).

13 Moreover, the same rheometer was used to carry out interfacial rheological measurements.
14 The geometry used was the Du Nouy ring (internal cup diameter 12.7 mm, external cup
15 diameter 25.6, internal ring diameter 19 mm, external ring diameter 25.4, volume 4 ml). The
16 shear rate was set between 0.1 and 100 s⁻¹. The system was positioned at the interface and
17 the sample was deliberately kept in direct contact with the ambient atmosphere. The soaking
18 time of the sample was 10 min.

19 *2.6 Microstructure and size of the particles*

20 The microstructure of the electro sprayed particles was determined using the SEM (JEOL JSM
21 6360) at an accelerating voltage of 20 kV and a working distance of 9-12 mm, equipped with
22 an image acquisition system. Samples were sputtered with a gold-palladium mixture under
23 vacuum before examining their structure. Several pictures were taken from random sample
24 positions representing the overall structure of each sample.

25 The particle size distributions were analyzed by Dynamic Laser Light Scattering equipment
26 (Zetasizer Nano ZS, Malvern Instruments, Worcestershire, UK) using water as dispersing
27 medium. Triplicate measurements were recorded at ambient temperature and the results are
28 expressed as the mean value. The polydispersity index is also evaluated using the following
29 equation:

30
$$PDI = \frac{D_{90} - D_{10}}{D_{50}} \quad (\text{Eq. 1}).$$

1

2 *2.7 Statistical analysis*

3 Statistical analysis of the results was performed with Statgraphics Centurion XV
4 (Statgraphics, Rockville, MD, USA) and an ANOVA test was applied in order to compare the
5 mean values of selected properties at a 95% confidence level. The same software was used
6 to perform clustering analysis, principal component analysis (PCA) and Pearson's multiple
7 variable analysis.

8

9 **3. Results and discussion**

10 *3.1 Physical properties of the BCNFs-WPI suspensions*

11 Existing work has shown that the physical properties of polymer suspensions such as
12 viscosity, surface tension and conductivity play a key role in the successful formation of
13 particles (Kriegel, McClements & Weiss, 2009). From Table 1, it can be observed that a
14 drastic increase in the surface tension occurs by increasing the BCNFS concentration of the
15 suspension. Specifically, for small amounts of BCNFS (1% wt) the surface tension bears
16 values of 20 mN/m, while for higher ones (16% wt BCNFS) the values go up to 40 mN/m,
17 regardless of the WPI concentration.

18

19 **Table 1.** Physical properties (surface tension, conductivity and ζ -potential) of suspensions
20 containing 1-16% wt BCNFs and 10-30% wt WPI and the polydispersity index (PDI) of the
21 produced particles

Physical Properties	%wt BC %wt WPI	1	2	4	8	16
		Surface tension (mN/m)	0	20.1 ^a (2.9)	27.3 ^b (2.4)	30.0 ^c (2.3)
Conductivity	10	20.6 ^a (0.7)	26.7 ^b (0.6)	30.1 ^c (0.5)	32.7 ^d (0.7)	38.6 ^f (1.2)
	20	20.2 ^a (1.7)	27.4 ^b (1.5)	30.1 ^c (1.5)	32.8 ^d (0.5)	40.5 ^f (1.2)
	30	20.2 ^a (1.1)	27.9 ^b (1.5)	31.1 ^c (1.7)	32.9 ^d (4.3)	40.7 ^f (0.8)
	0	1.0 ^a (0.1)	2.2 ^c (0.1)	2.5 ^d (0.0)	3.1 ^e (0.0)	3.3 ^f (0.1)

(mS/m)	10	1.3 ^b (0.0)	2.4 ^d (0.0)	2.5 ^d (0.0)	3.1 ^e (0.2)	3.3 ^f (0.1)
	20	1.4 ^b (0.1)	2.5 ^d (0.2)	2.9 ^e (0.1)	3.1 ^e (0.1)	3.3 ^f (0.0)
	30	1.4 ^b (0.0)	2.6 ^d (0.0)	2.9 ^e (0.2)	3.1 ^e (0.0)	4.0 ^g (0.2)
	0	-20.6 ^e (0.7)	-21.5 ^f (1.4)	-21.6 ^f (1.5)	-25.9 ^h (0.4)	-28.3 ⁱ (0.8)
ζ-potential (mV)	10	-18.3 ^c (0.9)	-19.7 ^d (0.3)	-20.5 ^e (0.6)	-21 ^e (0.4)	-23.1 ^j (1.0)
	20	-15.3 ^b (1.2)	-17.1 ^c (0.9)	-17.3 ^c (1.8)	-17.8 ^c (2.1)	-18.8 ^c (0.8)
	30	-11.9 ^a (0.7)	-12.6 ^a (1.4)	-14.1 ^b (1.3)	-14.6 ^b (0.4)	-15.9 ^b (0.7)
	0	-	-	-	-	-
PDI (-)	10	0.8 ^a (0.0)	1.0 ^b (0.0)	0.9 ^{ab} (0.0)	0.9 ^{ab} (0.0)	1.0 ^b (0.0)
	20	0.9 ^{ab} (0.0)	0.9 ^{ab} (0.0)	1.1 ^c (0.0)	1.7 ^d (0.1)	2.1 ^e (0.1)
	30	1.0 ^b (0.0)	1.0 ^c (0.0)	1.6 ^d (0.1)	2.6 ^f (0.2)	3.1 ^g (0.3)

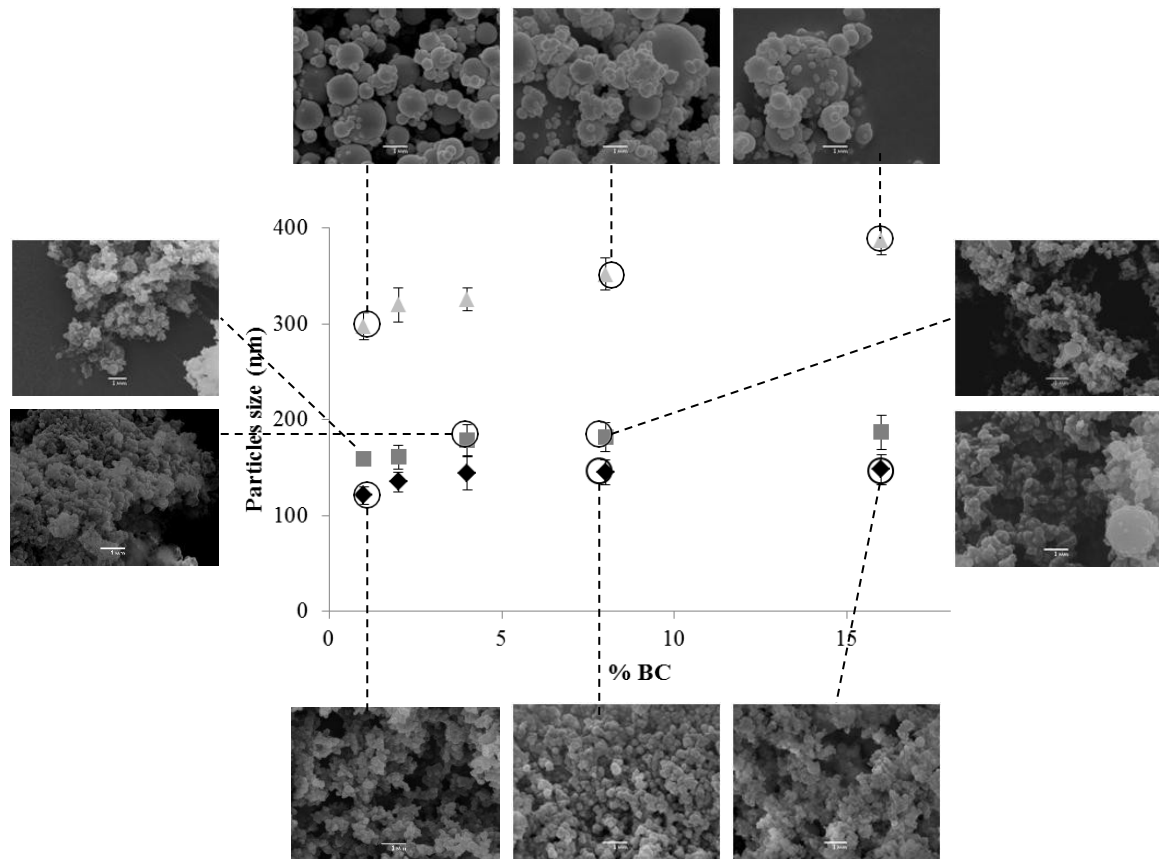
1 In parenthesis standard deviation values.

2 Mean values followed by the same letters are not significantly different (P > 0.05).

3

4 Hydrophilicity is connected to lower contact angle and higher surface tension. It is also
5 mentioned that BC fibrils can create many inter-and intra-fibrillar hydrogen bonds reducing the
6 fibrils dimensions and increasing hydrophilicity (Jia et al., 2016). According to Ganzevles
7 (2007) the presence of an excess of anionic polysaccharide can hinder the formation of a
8 dense protein layer at the air/water interface, justifying the similar values observed by
9 increasing WPI concentration.

10 Surface tension values are of high importance as they play a major role in the final size of the
11 particles. Furthermore, differences in the size of the processed system could be mainly
12 attributed to changes in the surface tension of the suspensions. Specifically, a decrease in
13 surface tension from 40 to 20 mN/m at suspensions containing 16 to 1% wt BCNFs and
14 constant amount of WPI (20% wt), leads to a decrease of the particle size from 390 to roughly
15 300 nm (Fig 1).



1

2 **Fig. 1.** Mean particle size (d_{50}) and microstructure (scale $1\mu\text{m}$) of the different BCNFS-WPI
 3 particles obtained through electrospaying of 1-16% wt BCNFs and 10% (\ominus), 20% (\circ), 30 (\square)
 4 wt % WPI suspensions.

5 This observation could have been due to the lower surface tension gradient around the
 6 droplet that causes the formed Taylor's cone to be expelled from the needle. It is found that
 7 the diameter of the particles could only be reduced with the stable cone-jet mode, which also
 8 improves the size of the particles. Moreover, Pancholi, Arhas, Stride & Ediringhe (2009)
 9 described a decrease of the mean particle size of chitosan particles, by decreasing the
 10 surface tension from 80 to 50 mN/m. A more hydrophobic surfactant was used for that
 11 purpose.

12 According to our previous study (Kaltsa, Paximada, Mandala & Scholten, 2014) pure WPI
 13 solution reach an interfacial pressure of 26 mN/m after a certain time. However, the surface
 14 tension of BCNFs-WPI suspensions does not significantly vary with the WPI concentration
 15 (Table 1)..

16 This result is attributed to the fact that BCNFs with their containing proteins move faster and
 17 cover the interface and then only a small amount of WPI is sufficient in saturating the surface.

1 When higher amounts of WPI are added to the system (20-30% wt) the excess protein
2 remains in the suspension and does not affect the surface tension. This is consistent with
3 other studies that worked with proteins and found no effect of the amaranth proteins on the
4 surface tension (Aceituno-Medina, Mendoza, Lagaron & Lopez-Rubio, 2013).

5 The conductivity values of the suspensions followed a steady increasing trend when the
6 BCNFS concentration increased throughout the whole experiment (Table 1). Specifically, by
7 increasing the BCNFS concentration from 1 to 16% wt, the electrical conductivity increased
8 from 1.3 to 3.3 mS/m, regardless of the WPI content. This phenomenon is attributed to the
9 predominant effect that the addition of higher BCNFs concentrations has to the system, as it
10 was noticed in surface tension values as well. In accordance to these results, Rošic, et al.
11 (2012) found an increase in conductivity by increasing polysaccharide concentration.

12 Conductivity is influenced by the polymer concentration because both natural polymers
13 display polyelectrolyte properties in aqueous media, i.e., WPI is positively charged in citrate
14 buffer and BCNFS is negatively charged in all the pH range.

15 The formation of a Taylor cone for electrospaying depends on the sufficiently high surface
16 charge densities. In turn, these densities depend on the conductivity of the biopolymer
17 suspension and the applied voltage (Yu, et al., 2011). Differences in the conductivity of
18 BCNFs are relatively small and do not explain the observed large changes in particles
19 diameter (Fig 1). On the other hand, the 30% wt WPI suspensions and high BCNFS amounts
20 that exhibit high conductivity values, led to a complicated process. It is worth noting that if
21 electrical conductivity is too high, the charge carried by the electrospaying jet is lower,
22 resulting in potential destabilization of the jet and the complication of the process (Bock,
23 Dargaville & Woodruff, 2012).

24 The ζ -potential representing the charge density of the BCNFs-WPI suspensions is
25 summarized in Table 1. It has been previously reported that BC bears a negative charge in all
26 the pH range (Paximada, Koutinas, Scholten & Mandala, 2016). Therefore, by increasing the
27 BCNFs concentration from 1 to 16% wt, the overall charge of the suspensions decreased
28 from -18 to -23 mV for suspensions containing 10% wt WPI. The same trend is also obvious
29 in suspensions containing 20 or 30% wt WPI.

1 A progressive increase of WPI concentration from 10 to 30%, at constant BCNFs amount,
2 leads in ζ -potential increase in all cases. It is known that, at low pH, since WPI is positively
3 charged, with ζ -potential equal to +35.9 mV in emulsions systems at Ph=3.0 according to
4 Kaltsa, Paximada, Mandala & Scholten (2014). Then the increase in ζ - potential is attributed to
5 protein and BCNFs associations. However, positive values were not observed. This may
6 happen because BC small fibrils, due to their large volume, determine the overall charge
7 density. Moreover, the rate of ζ -potential increase differs according to the initial BC amount. In
8 particular, at high BCNFs concentration, 8% or 16%, the change of ζ -potential is steeper. On
9 the contrary, at BCNFs concentration 1-4%, this rate is similar and lower. When the protein
10 amount is constant but BC increases, a slight decrease in the overall charge is observed.
11 Then the protein increase results in greater changes in ζ - potential than BC does, in
12 agreement to further results when particles characteristics are categorized according to
13 protein amount (cluster analysis, 3.4).

14 To conclude, BC has a dominant effect on the surface tension and conductivity, whereas WPI
15 influenced ζ -potential of the suspensions, leading to differences in the electrospraying
16 process and the produced particles.

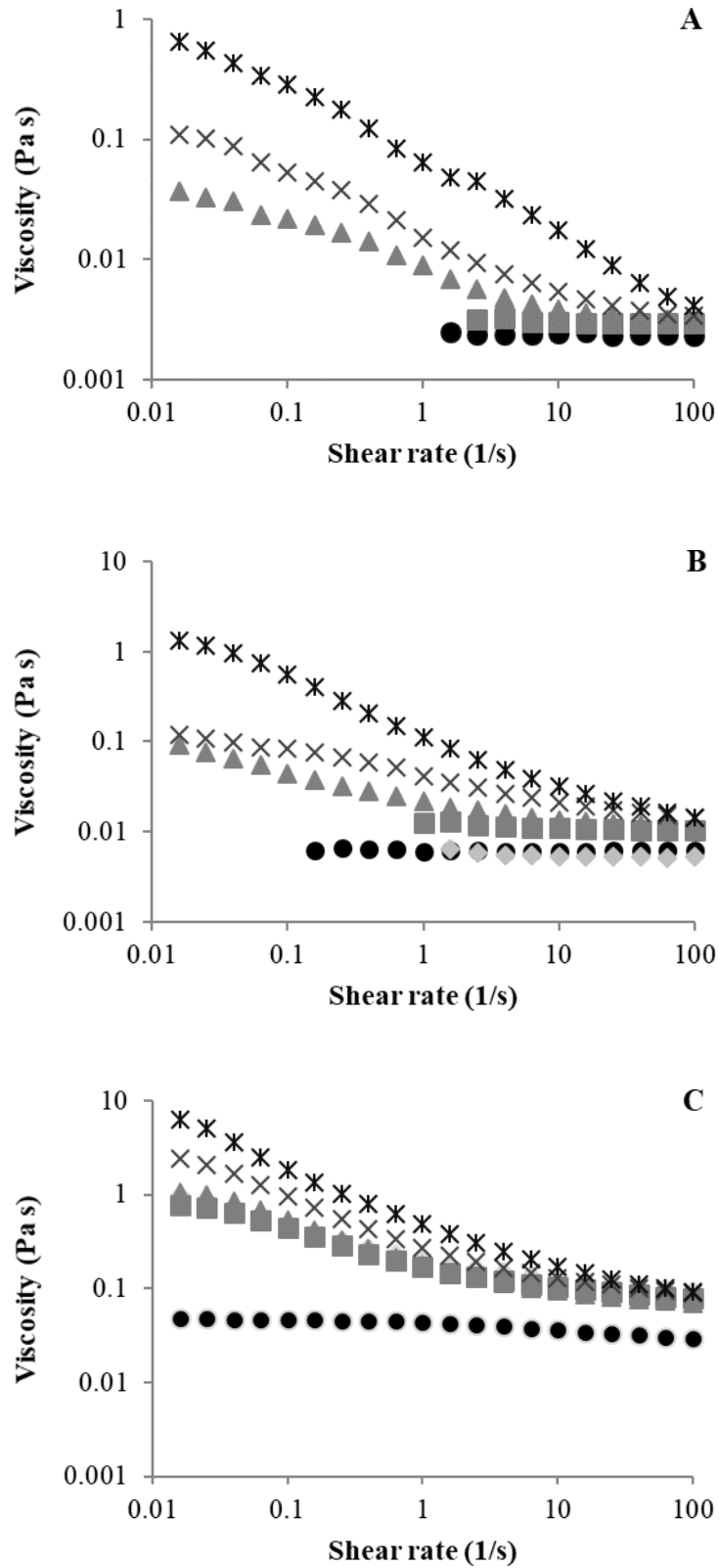
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18 *3.2 Bulk viscosity of the BCNFs-WPI suspensions*

19 The viscosity of the suspensions as a function of different BCNFs and WPI concentrations is
20 presented in Fig. 2 (A-C).

21

22



1 **Fig. 2.** Apparent viscosity curves of suspensions containing 0% wt (\circ), 1% wt (\triangle), 2% wt (\square),
 2 4% wt (\diamond), 8% wt (\times), 16% wt (\rightarrow) BCNFs and 10% (A), 20% (B), 30% (C) wt WPI.
 3

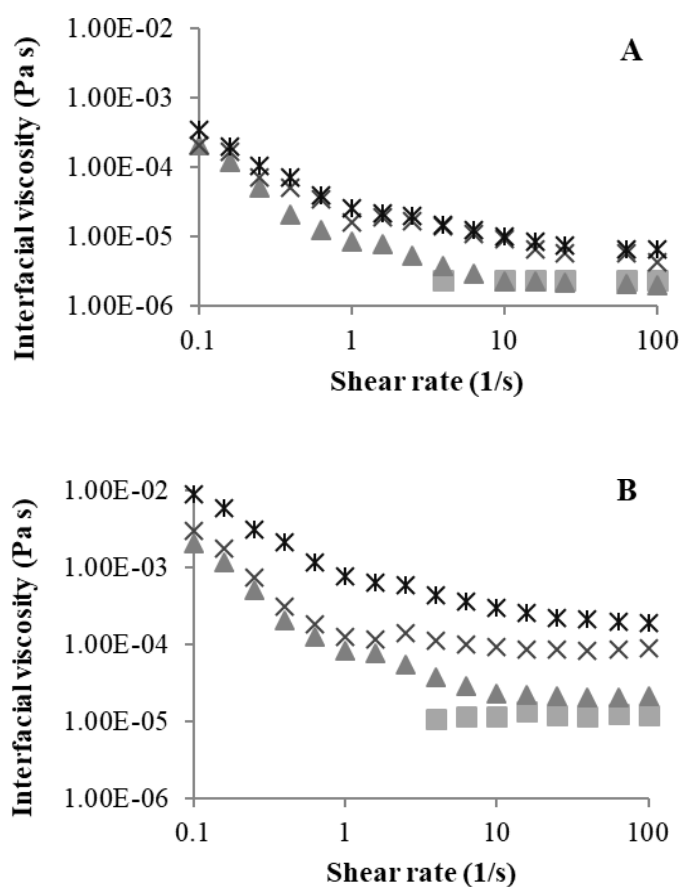
1 In the absence of BCNFs, all suspensions exhibit a Newtonian-like behavior as their viscosity
2 is constant in all shear rates, regardless of the WPI concentration. This Newtonian-like
3 behavior is typical for suspensions with a low WPI content (Paximada, Koutinas, Scholten &
4 Mandala, 2016). The viscosity of suspensions containing 10 or 20% wt and low amount of
5 BCNFs (1-4% wt) still exhibits Newtonian-like behavior, which is consistent with the idea that
6 in suspensions contain low amounts of biopolymers, WPI is the polymer that has the major
7 effect on the viscosity of the mixture. This has already been described from other studies in
8 the properties of protein and polysaccharide mixtures (Kaltsa, Paximada, Mandala &
9 Scholten, 2014).

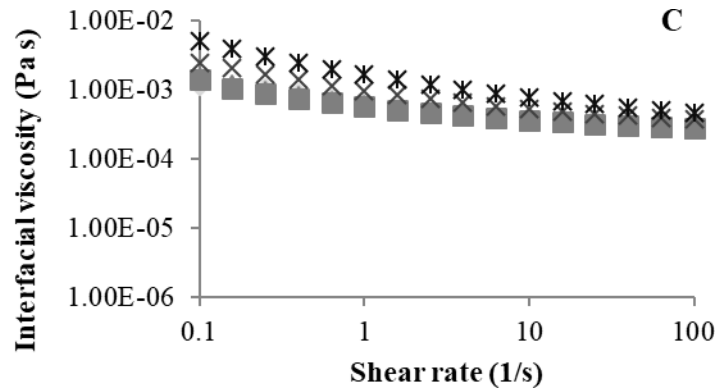
10 For the rest of the suspensions, their viscosity increases significantly from roughly 7mPas (no
11 BCNFS) to 0.7 -7 Pas, an increase of 2 to 3 orders of magnitude when BCNFS is from 1-16%.
12 The suspensions also exhibit shear thinning behavior, like most suspensions containing a
13 high concentration of polymers. A synergistic interaction between the anionic BCNFs and the
14 cationic WPI mixtures can be attributed either to the intermolecular binding that occurs
15 between side chains of the polymers, or to the interactions between disordered segments of
16 WPI molecules and BCNFs. This interaction enhances the viscosity of the suspensions
17 (Renou, Petibon, Malhiac & Grisel, 2013).

18 The pseudoplasticity of the suspensions increases with an increasing proportion of BCNFs in
19 the mixtures. The pronounced shear-thinning effect can be related to the structure of the
20 polymer chains in the suspensions. Before the beginning of the shearing, each
21 macromolecule has the shape of a three-dimensional coil as this is at the lowest energy state.
22 During the shear process, the molecules are tent to orient parallel to the direction of shear,
23 resulting in elongation, which lowers their flow resistance and results in a decrease in the bulk
24 viscosity (Freeman, 1996). Viscosity is one of the crucial parameters in predicting the ability of
25 one suspension to be electrospun or electrosprayed. High and low viscosities give rise to
26 dripping of suspensions, which is the case in the suspensions containing 30% WPI (Fig.1).
27 Studies that have examined different polymers on their ability to form particles, showed
28 viscosities at the range of 50-250 mPas for chitosan (Pancholi, Ahras, Stride & Edirisinghe,
29 2009), and 165 mPas for zein (Neo et al., 2013). At 16% BCNFS concentration and at 30%
30 WPI regardless of the BCNFS amount, far greater values were observed.

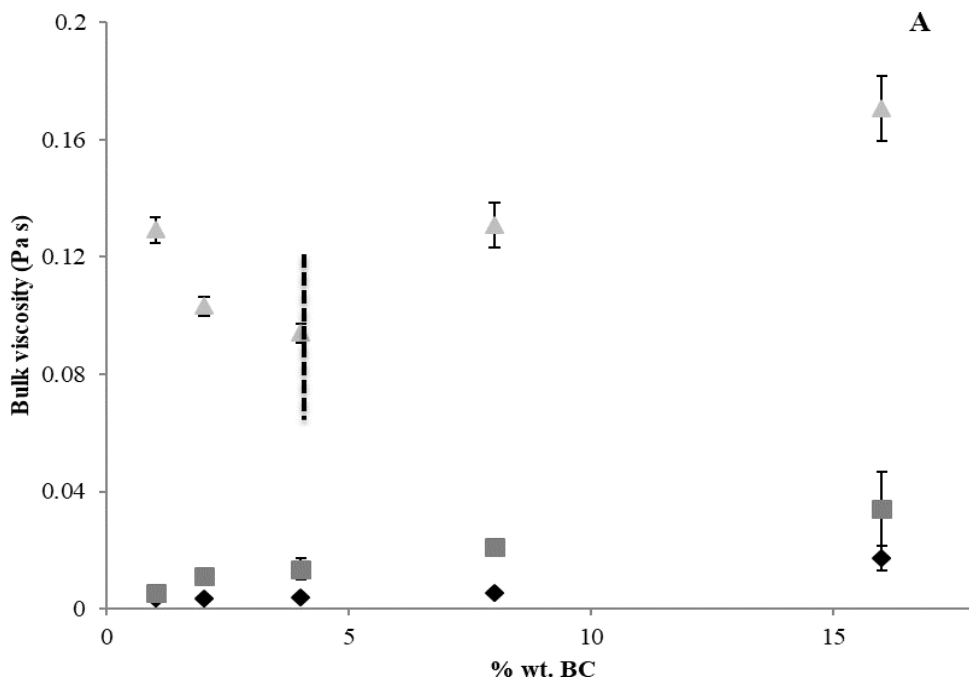
1 3.3 Interfacial viscosity of the BCNFS-WPI suspensions

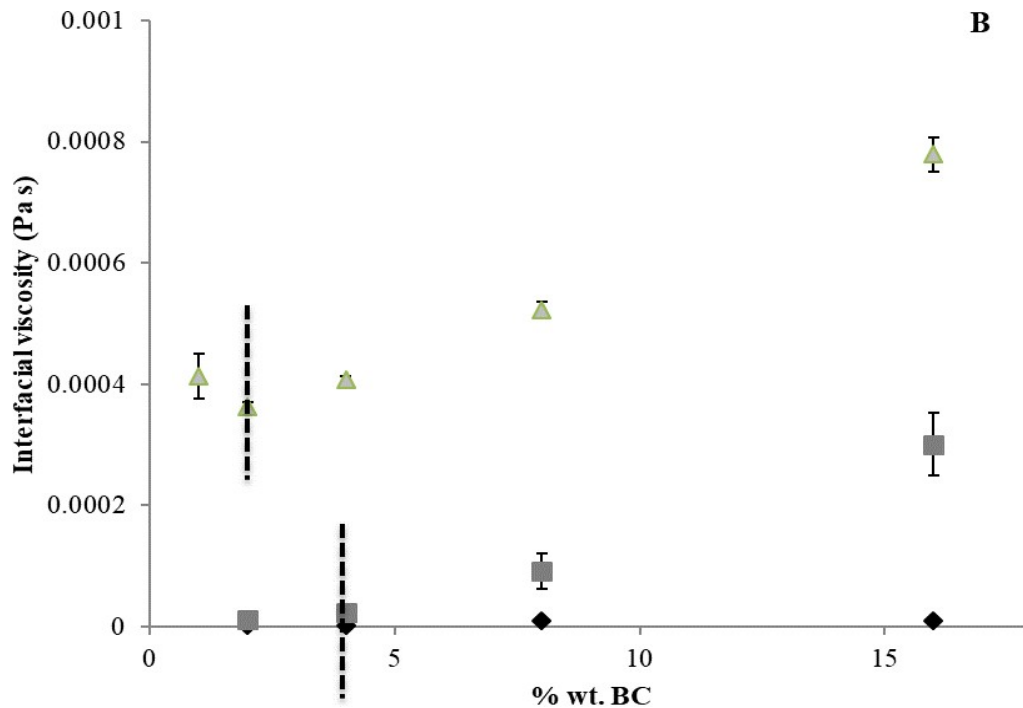
2 Numerous existing papers on electrosprayed or electrospun materials examined the viscosity
3 of the polymer suspensions (Aceituno-Medina, Mendoza, Lagaron & Lopez-Rubio, 2013;
4 Sarhan and Azzazy, 2015). Interfacial viscosity may bear an effect on the sprayability of the
5 polymers (Pelipenko et al., 2012) suggesting that, given the area of a Taylor cone, rheological
6 characteristics play a significant role due to the larger diameter of the jet. Hence,
7 measurement of the rheological parameters of the bulk may not be sufficient when dealing
8 with electrospraying. In this study the interfacial viscosity of the suspensions as a function of
9 different BCNFS-WPI concentrations is evaluated and summarized in Fig. 3 (A-C).





1 **Fig. 3.** Interfacial viscosity curves of suspensions containing 0% wt (∞), 1% wt (\oplus), 2% wt (\circ),
 2 4% wt (\square), 8% wt (\times), 16% wt (\rightarrow) BCNFS and 10% (A), 20% (B), 30% (C) wt WPI.
 3
 4 The results of the interfacial measurements presented in Fig. 4 show the same parameter
 5 trends as was observed in the bulk (Fig.3 A-C). However, the values of the interfacial
 6 viscosity are three to five orders of magnitude lower than those observed in the bulk, with the
 7 latter being comparable to data obtained by Rošic et al. (2012).





1 **Fig. 4.** Bulk (A) and interfacial viscosity curves (B) as a function of blended suspensions
 2 containing 1-16% wt BCNFs and 10% wt (○), 20% (◻) or 30% (◊) wt WPI. Bars indicating
 3 standard deviations.

4 Specifically, the viscosity of all suspensions containing BCNFS increases significantly from
 5 roughly 0.001mPas (1% BCNFS) to 5mPas (16% BCNFS) regardless of the amount of WPI
 6 that they contain. This increase in interfacial viscosity is attributed to the amount of BCNFS
 7 that is added to the system. Hence, BC acts as a thickener in the suspensions and its
 8 increase in both bulk and interfacial viscosity is in agreement with other studies that evaluated
 9 the thickening effect of BC (Panagopoulou, et al., 2015; Paximada et al., 2016a; Paximada et
 10 al., 2016b). Fig 4 illustrates the bulk and interfacial viscosity values (Fig.4A, 4B) as a function
 11 of the concentration of BCNFS in the suspensions. The values used for this graph were for
 12 both viscosities at shear rate of $10s^{-1}$. From this figure, it can be seen that, while for the
 13 suspensions containing 10% wt WPI the bulk and interfacial viscosity do not have significant
 14 differences, when the concentration of WPI is increased (20-30% wt) high differences are
 15 found. Specifically, the interfacial viscosity for the 20% wt WPI suspensions can be divided in
 16 2 regions: one from 1-4% wt BCNFs and the other from 4-16% wt BCNFs. As far as the
 17 suspension containing 30% wt WPI is concerned, the bulk viscosity shows a 2-region profile,

1 with the first region being from 1-4% wt and the second between 4-16% wt BCNFs. The
2 interfacial viscosity values of the first region were between 1-2% wt and between 2-16% wt of
3 BCNFs for the second.

4 A good correlation can be made between the viscosity values (Fig. 2A-C) and the
5 polydispersity index (PDI) of the particle's diameter (Table 1). In the case of 10% wt WPI, only
6 one region is depicted for both the bulk and interfacial viscosity values (Fig 5A, B), while the
7 PDI of all the produced samples was below 1 (Eq. 1). A PDI lower than 1, confirms the
8 monomodal size distribution of the particles (McClements, 2005). Another issue to consider is
9 the viscosity of the suspensions with 20% WPI. Only one region is observed for the bulk
10 viscosity (Fig. 4(A, B)). However, for the interfacial viscosity, 2 regions are observed, and they
11 are in agreement with the polydispersity of the suspensions which showed that from 1-4% wt
12 BCNFs the distribution is monomodal. However, by increasing the BCNFs amount, the
13 distribution becomes bimodal. Regarding the 2 regions observed for 30% wt WPI, the slope in
14 the first region includes suspensions having 4% wt or less BCNFs in the suspension, while
15 the second region includes BCNFs content of up to 16% wt. In contrast to the results of the
16 bulk viscosity, interfacial viscosity (Fig. 4B) exhibits 2 regions, the first being for BCNFs
17 concentrations up to 2% wt and the second up to 16% wt.

18 The interfacial viscosity values are in better agreement with the polydispersity of the produced
19 particles. Suspensions falling in the first region gave particles of monomodal size distribution
20 ($PDI < 1$), while those falling in the second region resulted in particles that exhibit a multimodal
21 distribution as their PDI was greater than 1.

22 Specifically, the absolute value of the correlation for bulk viscosity and PDI is lower (0.68)
23 than the values for interfacial viscosity and PDI (0.81), indicating a stronger linear relationship
24 between interfacial viscosity and PDI ($P < 0.05$).

25 In a previous study, the correlation between the rheological properties of the chitosan-PEO
26 blends and their electrospinnability can be seen (Rošić, et al., 2012). An interesting
27 observation was that interfacial viscosity gave better response in predicting the
28 electrospinnability of the suspensions. All in all, the correlation between the rheological
29 characteristics of the interface and nanofiber morphology is far more distinctive than the
30 correlations with rheological properties in the bulk.

1 3.4 Properties of the produced particles

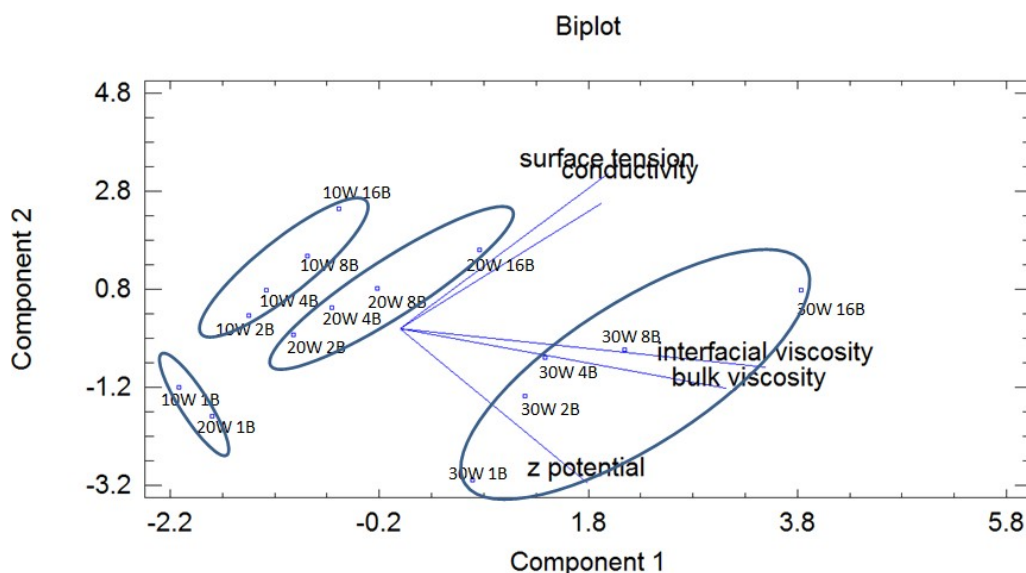
2 Fig. 4 shows SEM images of the structures obtained together with the mean particle diameter
3 and the polydispersity index (PDI) of the produced materials (Table 1). The mean diameter
4 (Z_{av}) of the produced particles was in the range of 120–390 nm for different compositions of
5 BCNFS-WPI. In general, the diameter of the electrosprayed particles depended on the
6 polymer concentration. An increase in both biopolymers causes an increase in the particle
7 diameter and PDI, in accordance with previously published data (Wongsasulak, McClements,
8 Yoovidhya & Weiss, 2007). For example, the Z_{av} of electrosprayed BCNFS-WPI particles
9 increased from 121 to 148 nm by increasing the amount of BCNFS from 1-16% wt and
10 keeping the WPI constant (10% wt).

11 Regarding the structure of the produced particles, the first observation is that particles with
12 low WPI (10 -20% wt) exhibit a more homogeneous structure. This could be explained by the
13 binding of BCNFS monomers to the WPI backbone either through hydrophobic interactions or
14 hydrogen bonding, forming a complex which affects intramolecular interactions and
15 conformation of the protein. As previously hypothesized, interaction between BCNFS and
16 protein may result in a more open molecular structure which may help to establish
17 interactions with the WPI, decreasing the critical entanglement concentration and facilitating
18 electrospraying (Kriegel, McClements & Weiss, 2009).

19 The addition of higher WPI amount (30% wt) leads to the particles with multimodal size
20 distributions and larger PDI values up to 3 (Table 1). Amorphous parts that are obvious in the
21 micrographs of these materials are caused by the dripping of the BCNFS-WPI suspensions
22 during the process. Albumins and globulins, the main components of WPI, have a globular
23 structure which cause inter- and intra-molecular interactions (McClements, 2005). The
24 mentioned α -helical and β -sheet polymer chain configurations which may be adopted in
25 higher WPI concentrations, also keep the rigidity of the chains, preventing the formation of
26 monodispersed particles through electrospraying.

27 Principal component analysis (PCA) is conducted in order to evaluate the properties that
28 mostly affect the size and structure of the particles (Fig. 5). In our case, 2 components have
29 been extracted, since they have eigenvalues greater than or equal to 1.0. Together they
30 account for 96.1% of the variability in our samples. As it can be seen, surface tension and

1 conductivity are mostly affected by BCNFs, as their eigenvalues are parallel to the increase of
 2 the BCNFs. This is in accordance with our previous measurements (3.1). On the other hand,
 3 bulk and interfacial viscosity are weighted more heavily in a positive direction by WPI
 4 concentration, as their eigenvalues are parallel to the augmentation of WPI. Another issue is
 5 that the variables furthest from 0, make the largest contribution to the system, which in our
 6 case are interfacial viscosity and surface tension. Only with these properties the structure of
 7 the electrospayed particles could be successfully analyzed. Furthermore, a cluster analysis is
 8 performed in order to categorize the samples (Fig. 5).
 9



10

11

Fig. 5. Principal component analysis of the electrospayed particles.

12

13

14 This procedure has created 4 clusters from the 15 observations supplied. Samples with 30%
 15 WPI comprise the first cluster, while the samples 10%WPI-1%BCNFs and 20%WPI-
 16 1%BCNFs the second. The samples with 10% WPI and 2-16% BCNFs comprise the third
 17 cluster, and the samples with 20% WPI and 2-16% BCNFs the fourth cluster. This
 18 categorization indicates that WPI plays a key role on grouping the samples, especially in the
 19 first cluster, which leads to dripping suspensions. It is obvious that BCNFs in low
 20 concentrations also plays an important role in grouping (cluster 2). Cluster 3 also exhibits low

1 viscosity and surface tension values, while cluster four (20% WPI, 2-16% BCNFs) yields
2 particles with the best properties.

3 In all cases, it was possible to produce particles with size distribution in the submicron range,
4 which is very promising as a first step of the encapsulation of bioactive compounds in the
5 BCNFS-WPI particles.

6

7 **4. Conclusions**

8 In this study, suspensions containing whey protein isolate (WPI) and bacterial cellulose (BC)
9 in various concentrations (1-16% wt BC and 10-30% wt WPI) were prepared and screened for
10 their electrosprayability. Surface tension and electrical conductivity were found to increase as
11 the BCNFs concentration increased, while they remained unaffected in terms of the protein
12 concentration. The increase in surface tension and conductivity with increasing BCNFS
13 concentration may be attributed to a change in the bonds that are formed between the
14 polysaccharide and the proteins. Viscosity and interfacial viscosity increased as the
15 proportion of BCNFS increased. A good correlation (0.83) was found between the interfacial
16 rheology values of the suspension and the distribution of the produced particles. The
17 produced particles varied in size (120 to 380 nm) and polydispersity (PDI= 0.8-3). After a PCA
18 analysis, it can be said that the properties that could best predict the structure of the
19 electrosprayed particles, are interfacial viscosity and surface tension. The suspension that
20 could be suggested as a good alternative material for microencapsulation is the one
21 containing 8% BCNFS and 20% WPI. All in all, the results of this study are encouraging as a
22 first step for the encapsulation of bioactive and volatile compounds in the BCNFs-WPI
23 particles.

24

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3

4 References

- 5 Aceituno-Medina, M., Mendoza, S., Lagaron, J. M., López-Rubio, A. 2013. Development and
6 characterization of food-grade electrospun fibers from amaranth protein and
7 pullulan blends. *Food Research International*, 54(1), 667-674.
- 8 Azeredo, H.M.C. 2018 Bacterial cellulose for food applications-A review. *International*
9 *Journal in Advances in medical biotechnology*, 1(2), 1-5
- 10 Bhardwaj, N., Kundu, S. C. 2010. Electrospinning: A fascinating fiber fabrication technique.
11 *Biotechnology Advances*, 28(3), 325-347.
- 12 Bock, N., Dargaville, T. R., Woodruff, M. A. 2012. Electro spraying of polymers with
13 therapeutic molecules: State of the art. *Progress in Polymer Science*, 37(11), 1510-
14 1551.
- 15 Casper, C. L., Stephens, J. S., Tassi, N. G., Chase, D. B., Rabolt, J. F. 2004. Controlling Surface
16 Morphology of Electrospun Polystyrene Fibers: Effect of Humidity and Molecular
17 Weight in the Electrospinning Process. *Macromolecules*, 37(2), 573-578.
- 18 Choi S.M. & Shin E.J. (2020) The Nanofication and Functionalization of Bacterial Cellulose
19 and Its Applications. *Nanomaterials*, 10, 406; 1-24.
- 20 Freeman, J. 1996. *Rheological Methods in Food Process Engineering*. Freeman Press, 418.
- 21 Giroux, H. J., Robitaille, G., Britten, M. 2016. Controlled release of casein-derived peptides in
22 the gastrointestinal environment by encapsulation in water-in-oil-in-water double
23 emulsions. *LWT - Food Science and Technology*, 69, 225-232.
- 24 Gouzy, R., Tsekou, C., Remiln, C., Velikov, K.P. 2019. Cellulose microfibril networks in
25 hydrolysed soy protein isolate solutions. *Colloids and Surfaces A: Physicochemical*
26 *and Engineering Aspects*, 568, 277-283.
- 27 Hsieh, Y.-C., Yano, H., Nogi, M., Eichhorn, S.-J. 2008. An estimation of the Young's modulus of
28 bacterial cellulose filaments. *Cellulose*, 15, 507-513.
- 29 Jia Y., Zhai X., Fu W., Liu Y., Li F., Zhong C., 2016, Surfactant-free emulsions stabilized by
30 tempo-oxidized bacterial cellulose *Carbohydrate Polymers*, 151 (20), 907-915.
- 31 Kaltsa, O., Paximada, P., Mandala, I., Scholten, E. 2014. Physical characteristics of submicron
32 emulsions upon partial displacement of whey protein by a small molecular weight
33 surfactant and pectin addition. *Food Research International*, 66(0), 401-408.
- 34 Koupantsis, T., Pavlidou, E., Paraskevopoulou, A. 2016. Glycerol and tannic acid as applied in
35 the preparation of milk proteins – CMC complex coavervates for flavour
36 encapsulation. *Food Hydrocolloids*, 57, 62-71.
- 37 Kriegel, C., Kit, K. M., McClements, D. J., Weiss, J. 2009. Electrospinning of chitosan–
38 poly(ethylene oxide) blend nanofibers in the presence of micellar surfactant
39 solutions. *Polymer*, 50(1), 189-200.
- 40 Li, Q., Wang, Y., Wu, Y., He, K., Li, Y., Luo, X., Li, B., Wang, C., Liu, S. 2019. Flexible cellulose
41 nanofibrils as novel Pickering stabilizers: The emulsifying property and packing
42 behavior. *Food Hydrocolloids*, 88, 180-189.
- 43 McClements, D. J. 2005. *Food Emulsions: Principles, Practice and Techniques*. Boca Raton,
44 CRC Press.
- 45 Neo, Y. P., Ray, S., Jin, J., Gizdavic-Nikolaidis, M., Nieuwoudt, M. K., Liu, D., Quek, S. Y. 2013.
46 Encapsulation of food grade antioxidant in natural biopolymer by electrospinning
47 technique: a physicochemical study based on zein-gallic acid system. *Food Chem*,
48 136(2), 1013-1021.

- 1 Nishi, Y., Uryu, M., Yamanaka, S., Watanabe, K., Kitamura, N., Iguchi, M. 1990. The structure
2 and mechanical properties of sheets prepared from bacterial cellulose. *Journal of*
3 *Materials Science*, 25 (6), 2997-3001.
- 4 Panagopoulou, E., Tsouko, E., Kopsahelis, N., Koutinas, A., Mandala, I., Evageliou, V. 2015.
5 Olive oil emulsions formed by catastrophic phase inversion using bacterial cellulose
6 and whey protein isolate. *Colloids and Surfaces A: Physicochemical and Engineering*
7 *Aspects*, 486, 203-210.
- 8 Pancholi, K., Ahras, N., Stride, E., Edirisinghe, M. 2009. Novel electrohydrodynamic
9 preparation of porous chitosan particles for drug delivery. *J Mater Sci Mater Med*,
10 20(4), 917-923.
- 11 Paximada, P., Dimitrakopoulou, E. A., Tsouko, E., Koutinas, A. A., Fasseas, C., Mandala, I. G.
12 2016a. Structural modification of Bacterial Cellulose fibrils under ultrasonic
13 irradiation. *Carbohydrate Polymers*.
- 14 Paximada, P., Koutinas, A. A., Scholten, E., Mandala, I. G. 2016b. Effect of bacterial cellulose
15 addition on physical properties of WPI emulsions. Comparison with common
16 thickeners. *Food Hydrocolloids*, 54, Part B, 245-254.
- 17 Paximada, P., Tsouko, E., Kopsahelis, N., Koutinas, A. A., Mandala, I. 2016c. Bacterial
18 cellulose as stabilizer of o/w emulsions. *Food Hydrocolloids*, 53, 225-232.
- 19 Paximada, P., Dubey, B., Howarth, M. 2018. Electrospayed particles from nano-emulsions as
20 carriers of fish oil. *Techconnect Briefs*, 1-4.
- 21 Pelipenko, J., Kristl, J., Rosic, R., Baumgartner, S., Kocbek, P. 2012. Interfacial rheology: an
22 overview of measuring techniques and its role in dispersions and electrospinning.
23 *Acta Pharm*, 62(2), 123-140.
- 24 Regev, O., Vandebril, S., Zussman, E., Clasen, C. 2010. The role of interfacial viscoelasticity in
25 the stabilization of an electrospun jet. *Polymer*, 51(12), 2611-2620.
- 26 Renou, F., Petibon, O., Malhiac, C., Grisel, M. 2013. Effect of xanthan structure on its
27 interaction with locust bean gum: Toward prediction of rheological properties. *Food*
28 *Hydrocolloids*, 32(2), 331-340.
- 29 Rošic, R., Pelipenko, J., Kocbek, P., Baumgartner, S., Bešter-Rogač, M., Kristl, J. 2012. The role
30 of rheology of polymer solutions in predicting nanofiber formation by
31 electrospinning. *European Polymer Journal*, 48(8), 1374-1384.
- 32 Sarhan, W. A., & Azzazy, H. M. E. 2015. High concentration honey chitosan electrospun
33 nanofibers: Biocompatibility and antibacterial effects. *Carbohydrate Polymers*, 122,
34 135-143.
- 35 Theron, S. A., Zussman, E., Yarin, A. L. 2004. Experimental investigation of the governing
36 parameters in the electrospinning of polymer solutions. *Polymer*, 45(6), 2017-2030.
- 37 Winuprasith, T., Suphantharika, M. (2013). Microfibrillated cellulose from mangosteen
38 (*Garcinia mangostana* L.) rind: Preparation, characterization, and evaluation as an
39 emulsion stabilizer. *Food Hydrocolloids*, 32(2), 383-394.
- 40 Wongsasulak, S., Kit, K. M., McClements, D. J., Yoovidhya, T., Weiss, J. 2007. The effect of
41 solution properties on the morphology of ultrafine electrospun egg albumen-PEO
42 composite fibers. *Polymer*, 48(2), 448-457.
- 43 Yu, D.-G., Branford-White, C., Williams, G. R., Bligh, S. W. A., White, K., Zhu, L.-M.,
44 Chatterton, N. P. 2011. Self-assembled liposomes from amphiphilic electrospun
45 nanofibers. *Soft Matter*, 7(18), 8239-8247.
- 46 Zhang, X., Liu, Y., Wang, Y., Luo, X., Li, Y. Li, B. 2019. Surface modification of cellulose
47 nanofibrils with protein nanoparticles for enhancing the stabilization of O/W
48 pickering emulsions. *Food Hydrocolloids*, 97, 105-180.
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*Credit Author Statement

Dr. P. Paximada was responsible for conceptualization, data curation, investigation , and writing the original draft, Mrs E. Kanavou was responsible for data curation and Dr. I. Mandala was responsible for conceptualization, supervision, writing-review & editing